

Chronic Obstructive Pulmonary Disease: Effective Treatment with Vasoactive Intestinal Peptide (VIP)

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Abstract

Rationale: Chronic obstructive pulmonary disease (COPD) is characterized by progressive airflow limitation and chronic inflammation including the increased expression of Interleukin 8 (IL-8) and tumor necrosis factor (TNF, previously known as tumor necrosis factor alpha). We describe a novel physiological role of the Vasoactive Intestinal Peptide (VIP) and its therapeutical potential in COPD. Our rationale is based on a deficiency of VIP in the serum of COPD patients.

Objectives: The aim of this study was to evaluate a new therapy of COPD.

Methods: Molecular biological and immunochemical methods were used to characterize the action of VIP at the cellular level. A double-blind, placebo-controlled, randomized study was undertaken to evaluate the potential clinical role of VIP in COPD.

Measurements and Main Results: VIP treatment downregulates the expression of IL-8 and TNF that coincides with the inhibition of transcription factors AP-1, Stat3 and NF-κB. VIP treatment improves lung function and prognostic parameters of COPD. In a clinical study with 34 COPD patients, forced expiratory volume in 1s significantly improved with VIP by 0.107l and vital capacity by 0.155l compared with placebo (P<0.01).

According to the short 36-item questionnaire and to St. George's respiratory questionnaire, the improvement in quality of life was greater with VIP (4.08; -5.25) than with placebo (0.20; 1.72). Similarly, exercise capacity increased by 24.1 m (P<0.05) in the VIP group as compared to placebo (-8.6m) using the six minutes walking test.

Conclusions: This study is the first to show the immunoregulatory effect of VIP in COPD patients, and supports the notion of inhaled VIP as an attractive future therapy to dampen exaggerated immune responses in chronic lung disorders. Our data provide "proof of concept" for further investigation of VIP and its potential role in COPD.

Keywords: COPD; chronic inflammation; lung biopsy; vasoactive intestinal peptide

1. Introduction

Chronic obstructive pulmonary disease (COPD) is the third leading cause of death worldwide, causing 3.4 million deaths in 2023, approximately 6% of all global deaths. COPD is the overall term for a group of chronic conditions that are associated with the obstruction of lungs' airways, usually referring to the following disorders: Chronic bronchitis, Bronchiectasis and Emphysema. The most common symptoms of COPD include shortness of breath, chronic coughing, chest tightness, greater effort to breathe, increased mucus production and frequent clearing of the throat.

The mainstay medication consists of anticholinergic and beta-adrenergic bronchodilators. Addition of corticosteroid therapy may enhance bronchodilator responses and reduce exacerbations. An important feature of COPD is the ongoing chronic inflammatory process in the airways as indicated by the current GOLD (Global Initiative for Chronic Obstructive Lung Disease) definition. Neutrophil granulocytes, macrophages, and T-cells are involved in the cellular mechanisms of inflammation in COPD and are associated with an increased release of pro- and anti-inflammatory mediators such as e.g. IL-8 and TNF, a protease-antiprotease imbalance, oxidative stress, viral infections, and a genetic predisposition (e.g. Alpha-1 Antitrypsin Deficiency) [8, 13, 14, 23, 26, 30, 34, 35, 36, 39, 40, 42, 43, 45, 46].

We were looking for a drug that provides the combined potential advantages of lowering inflammation of the lung tissues, lowering pulmonary arterial pressure, reducing bronchoconstriction, improving blood circulation to the heart and lung, enhancing healing of bronchial epithelial cells, and improving the cardiovascular deficiencies in COPD.

Vasoactive Intestinal Peptide (VIP) is an abundant biologically active peptide endogenous in humans as well as in other mammalian species. It is produced by neurons in the peripheral and central nervous system, by endocrine cells, T-lymphocytes, and B-lymphocytes. This natural peptide is one of the signal molecules of the neuroendocrine-immune network comprising vasodilating, anti-proliferative, anti-inflammatory, and immune-regulatory features. Its predominant biological activity is performed in the lungs, and a vast body of experimental, pharmacological as well as clinical evidence suggests VIP to be an attractive candidate for the treatment of COPD [3, 9, 11, 12, 16, 18, 53, 54].

VIP's biological effects are mediated through the drug's pharmacological activity on VPAC1, VPAC2, and PAC-1 receptors (7-membrane-spanning G-protein coupled receptors) at the transcriptional level via modulation of NF- κ B, AP-1, Stat3, and cAMP responsive element (CRE)-binding, or ets-2 complexes in target cells [5, 12, 50]. The presence of VPAC receptors has been shown on airway epithelial cells, on macrophages surrounding capillaries and in the subintima of pulmonary arteries and veins [5, 50]. Stimulation of VPAC receptors activates pathways which have been shown to mediate the action of prostacyclins, nitric oxide, and phosphor diesterase inhibitors [19, 52].

We and others [21], noted that in patients with the lowest VIP serum levels, a more rapid formation of COPD was observed. The smallest values of VIP and the greatest changes in cardiopulmonary parameters were found in the cohort of COPD-patients with concomitant hypertension [21].

As a preferred route of administration, mouth inhalation was chosen. Previously we showed that after intravenous administration VIP is rapidly

distributed to the lungs, where it gets completely metabolized into biologically inactive parts by neutral endopeptidase, mast cell tryptase and mast cell chymase [37]. In general, inhaled drugs intended for lungs act quickly, minimize undesired negative side effects, avoid the hepatic first-pass metabolism, and act locally. As the size variability among adult lungs is smaller than the overall body size variability, the dosing reliability is also improved when inhaling.

We and others have shown that inhaled VIP decreases pulmonary artery pressure and pulmonary vascular resistance in patients with pulmonary hypertension, and exerts immunoregulatory and anti-inflammatory effects in sarcoidosis [37, 59, 60]. However, the exact mode of action of VIP is not yet completely understood.

After producing cigarette smoke-induced COPD in a mouse model, and after treatment of those mice with VIP, the levels of antioxidant-related factors (malondialdehyde (MDA) and superoxide dismutase (SOD)), fibrotic factors (hydroxyproline (HP) and TGF- β), pro-inflammatory cytokines (TNF, IL-1 β , and IL-6), and inflammation were diminished and controlled by VIP. The anti-inflammatory, anti-fibrotic, and anti-oxidant properties of VIP may be effective therapy in COPD [6].

Furthermore, the anti-inflammatory properties of IK312532 respirable powder (RP) - a stabilized VIP derivative - were characterized in an asthma/COPD-like animal model. Marked inflammatory events in the lung were observed after OVA-RP challenge in rats as evidenced by significant increase of inflammatory biomarkers such as eosinophil peroxidase (EPO), myeloperoxidase (MPO) and lactate dehydrogenase (LDH). However, intratracheal administration of IK312532-RP led to significant attenuation of plasma EPO, MPO and LDH activities, as well as significant reduction of recruited inflammatory cells in bronchoalveolar lavage fluid, especially macrophages and eosinophils. Thus, inhalable powder formulation of IK312532 exerts its anti-inflammatory activity by suppressing granulocyte recruitment to the lung and epithelial hyperplasia, followed by the reduction of cytotoxic peroxidases [29].

Alveolar macrophages (AM) are key players in the pathogenesis of COPD and contribute to the severity and progression of the disease. AM from COPD patients show a strong VPAC1 expression which exceeds VPAC2 expression. A similar receptor expression pattern was also observed in lipopolysaccharide (LPS)-activated monocyte-derived macrophages (MDM) from healthy volunteers and COPD patients. VIP has been shown to down-regulate IL-8 secretion significantly in MDM after LPS stimulation. The response to VIP was similar in MDM from COPD patients and healthy volunteers. Our previous results indicate that VPAC1 up-regulation in macrophages is a common mechanism in response to acute and chronic pro-inflammatory stimuli. Although VPAC1 up-regulation is dominant, both receptor subtypes are necessary for optimal anti-inflammatory signaling. The high VPAC1 expression in AM may reflect the chronic pro-inflammatory environment found in the lung of COPD patients. Treatment with VIP may help to decrease the chronic inflammation in the lung of COPD patients [4].

In view of what is known about VIP we correlated the serum concentration of VIP with the disease state of COPD and examined some of its effects on the transcription and expression of two essential cytokines in COPD both in vitro and in vivo. Moreover, we identified the essential therapeutic role of VIP in COPD patients in a small double-blind placebo controlled clinical trial.

2. Materials and Methods

VIP serum concentration

VIP was determined by radioimmunoassay (123I-VIP RIA) in healthy controls n=48, COPD patients (GOLD II) n=12, COPD (GOLD III) n=16, COPD (GOLD IV) n=14 [37].

Cell isolation and characterization

We cultured lung fibroblasts and bronchial smooth muscle cells (BSMC). All primary cell lines were isolated from resected lung biopsies of COPD patients undergoing therapeutic surgery for other reasons, after agreement of the local ethical committee and written informed consent of the patients. Human lung fibroblasts were isolated from bronchial biopsies and cultured as described earlier [48].

BSMC were isolated and grown from resected bronchi of the above patients as previously described, and BSMC were characterized by positive immuno-staining for α -smooth muscle actin and calponin [20].

Cell treatment

Sub-confluent (80%) cell cultures were serum deprived for 24 hours in medium without serum. The cells were then pre-incubated for 30 minutes with VIP (10⁻⁶ M - 10⁻¹² M) and then stimulated with the respective growth medium with VIP added at various concentrations.

Total RNA extraction and RT-PCR and Real-Time RCR-analysis mRNA was obtained from biopsy by TRIZOL using standard protocols and was directly transcribed into c-DNA using a reverse transcription kit (Advantage RT-for-PCR Kit) according to instructions of the manufacturer and was stored at minus 80°C until analysis.

Relative mRNA expression was determined by the ACT method as previously described [25]. The ACT numbers indicate the difference in amplification cycles between target and housekeeping (18s) gene.

Immunoblotting

Nuclear/cytosolic proteins were prepared with NE-PER kit according to the manufacturer protocol. Protein extracts were then size- fractionated by SDS-PAGE electrophoresis and transferred onto nitrocellulose membranes as described previously [41] and protein transfer was confirmed by Ponceau staining.

Electrophoretic mobility shift assay

Nuclear protein extracts were prepared after 30 min by NE-PER extraction kit and the protein content was determined by Bradford assay. The oligonucleotide was labeled using [Y33P]-dATP (3,000 Ci/mmol) and a T4 polynucleotide kinase. Samples were size fractionated by electrophoresis in a non-denaturing 4% polyacrylamide gel [41].

Cytokine enzyme linked immuno sorbent assay (ELISA)

Cytokines (IL-8 and TNF) were quantified in cell culture medium samples collected at below indicated time points from the same cell culture vessel for each cell line. Commercially available ELISA for IL-8, and TNF were used following the instructions of the manufacturer.

Immunohistochemistry for IL-8 and TNF

Rehydrated paraffin sections were stained for IL-8 and TNF according to the procedures as described previously [37].

Clinical study

This parallel-group, double-blind, placebo-controlled, randomized, monocenter study was undertaken at the Medical University of Vienna, Austria. The clinical study was cleared by the positive ethics approval from the Ethics Committee of the Medical University of Vienna, reference number 168/2003 and announced by the ClinicalTrials.gov ID NCT00464932.

The randomization sequence was generated by Rancode in a blinded manner; no person involved in data analysis had knowledge of the randomization sequence. Treatment was assigned by the investigators with sequential study numbers according to a block randomization list in ratios of 1:1 (VIP:placebo). VIP was synthetically manufactured. It was applied by a 10-minute inhalation (50µg VIP diluted in 3 ml of 0.9% NaCl solution), four-times daily, using a mesh nebulizer providing an average mass median aerodynamic diameter (MMAD) VIP particle size of 3µm to allow for a deep-lung deposition of the drug.

The trial substance was an “add-on” treatment; concomitant respiratory medications allowed throughout the study were short- and long-acting β -mimetics, long-acting anticholinergics, inhaled corticosteroids and theophyllines at a constant daily dose. Patients could receive a course of oral corticosteroids for treatment of exacerbations during the treatment phase.

The primary outcome variables were exercise capacity measured using the six-minute walking test (6MWT), Borg dyspnea score and health-related quality of life (HRQL) indicators measured by means of the short 36-item questionnaire (SF-36) developed for medical outcomes studies and St. George’s respiratory questionnaire (SGRQ). Findings were calculated as the change from base-line to the exit.

Secondary outcome measures were change from base-line in post-bronchodilator FEV1, post-bronchodilator inspiratory vital capacity (IVC) and number of COPD exacerbations. We recorded adverse events as part of the safety assessment using standard International Conference on Harmonisation Guidelines for Good Clinical Practice (ICH-GCP). Pulmonary function tests were done at rest before treatment onset (base-line), and at week 12 of the treatment period as recommended by the American Thoracic Society [44].

All patients were recruited from an outpatient setting. Inclusion criteria were: history of COPD >12 months, as defined by Global Initiative for chronic Obstructive Lung Disease (GOLD) [35], 40 years or older; current smoker or ex-smoker with a smoking history of >10 pack-years; post-bronchodilator FEV_i of 20-80% of predicted value; post-bronchodilator IVC ratio >70%; reversibility of FEV1 of <12% and/or <200 ml after 400 µg inhaled salbutamol; and stable clinical disease status with no change in COPD treatment during the last 4 weeks. Patients were excluded from study enrolment if they were diagnosed with asthma or other relevant lung diseases (e.g. lung cancer, bronchiectasis); if they had a recent exacerbation that required a course of systemic corticosteroids, emergency room treatment, or hospital admission within 4 weeks before the randomization. Patients were also excluded from study enrolment if they had a respiratory tract infection within 4 weeks before the randomization or known α 1-antitrypsin deficiency. The study was approved by the local ethical committee and each patient gave informed consent to participate in the trial and investigations according to institutional guidelines and to the Helsinki principles.

Statistical analysis

After completion of the case report forms, data capture was done with the double entry technique. Thereafter, the data were analyzed descriptively using the SPSS (IBM Statistics V.8) software package.

For analysis of the recruitment allocation, we compared means and standard deviation of relevant parameters (see table 1) of the patient groups.

Data analysis was done with SPSS, Version 12. Baseline demographic, lung function variables, quality of life are presented as means $\pm$ SD. Variables measured at baseline and during VIP inhalation in the same

patients were compared by Wilcoxon signed rank test (two-sided) for paired data. P values below 0.05 were considered statistically significant.

VIP serum levels were compared between the groups by means of the Students T-test, and by the Wilcoxon signed rank test when the data did not allow for computing the mean and standard deviations.

3. Results

We determined the serum concentration of VIP in humans by radioimmunoassay. As shown in Figure 1, patients with COPD show significantly reduced levels of VIP as compared to controls whereby the VIP levels correlated inversely with disease stage.

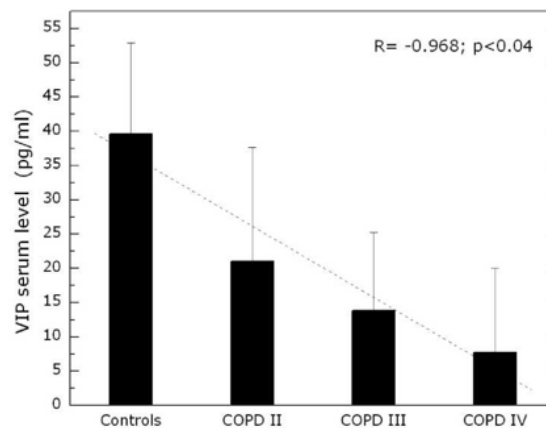
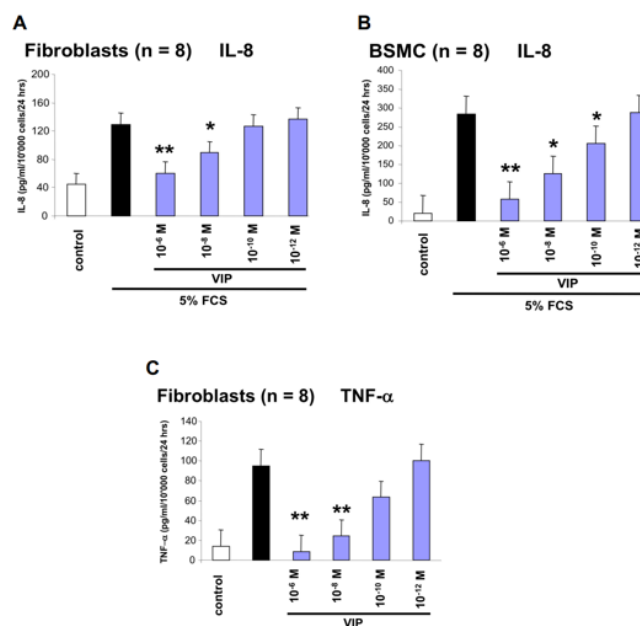


Figure 1: VIP serum concentration as determined by RIA of healthy controls and COPD patients. Healthy controls n=48, COPD (GOLD II) n=12, COPD (GOLD III) n=16, COPD (GOLD IV) n=14.

BSMC and fibroblasts obtained from COPD patients were stimulated with fetal calf serum (5%) following, the release of TNF and IL-8 was determined. Serum induced a significant release of both cytokines (Figure

2). This effect was counterbalanced by VIP as a function of concentration (IC₅₀, TNF 8.5×10^{-9} M and that for IL-8 2.6×10^{-9} M).



BSMC (n = 8) TNF

Bronchial smooth muscle cells did not produce significant levels of TNF within 24 hours.

Figure 2: Inhibitory effect of VIP on 5%FCS-stimulated IL-8 and TNF secretion by human lung BSMC and fibroblasts (A) IL-8 in BSMC (B) in fibroblasts (C) TNF in fibroblasts** indicates p Less-than sign 0.001, * indicates p Less-than sign 0.05 student's paired t-test compared to 5% FCS. Bars represent the mean $\pm$ S.E.M. of three independent experiments performed in four individual primary human BSMC lines or fibroblasts. BSMC did not secrete TNF under the described cell culture conditions within 24 hours.

Bronchial biopsies were used for examining the mRNA transcription and protein expression of the cytokines before and following the inhalation of VIP. The biopsies were taken from 6 patients at baseline and after 3 months of VIP inhalation (50 μ g, 4 x daily). Compared to 3 healthy controls RT-PCR results of biopsies obtained from COPD patients

showed an increased transcription of IL-8 and TNF before therapy at baseline. In contrast, as depicted in Figure 3, the transcription of both genes was down-regulated in the tissue samples from COPD patients following treatment with VIP.

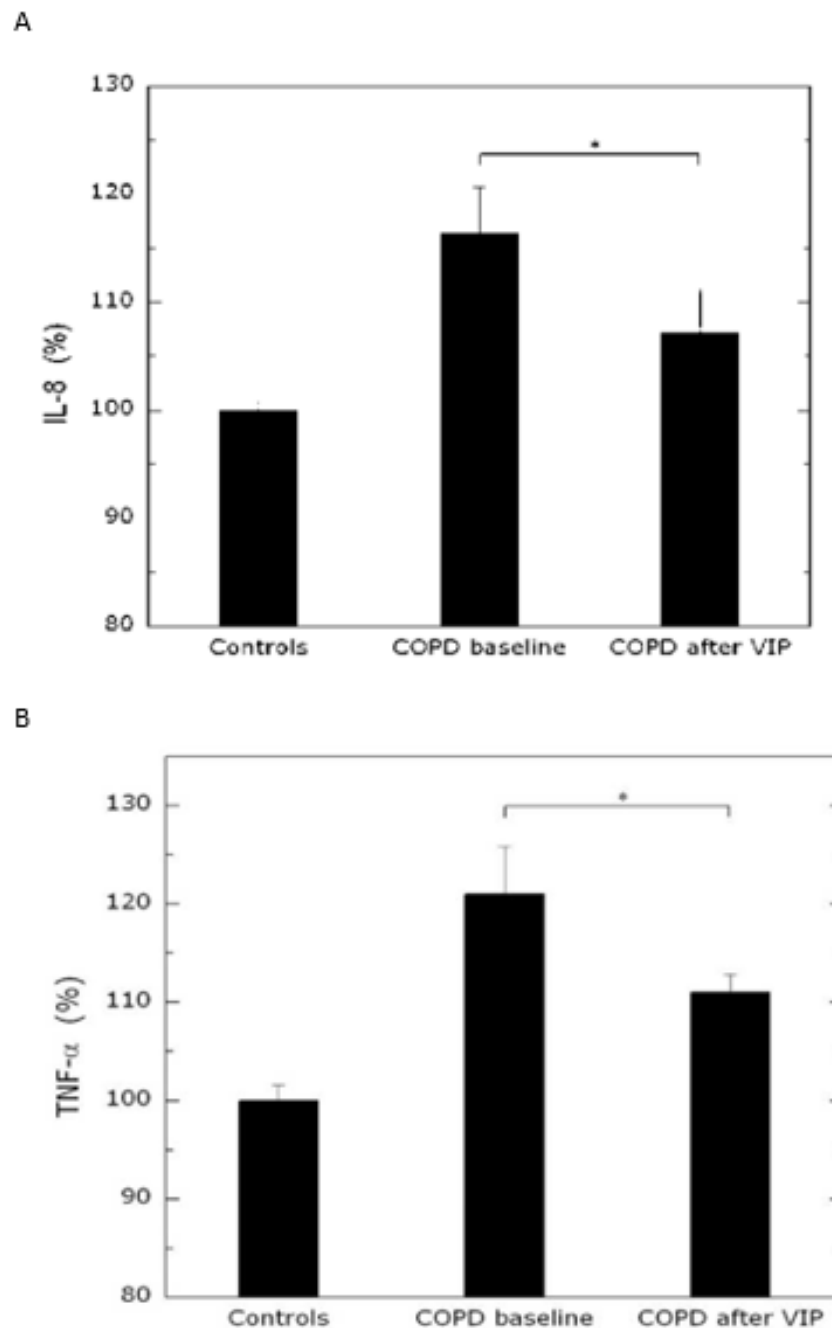


Figure 3: Transcription of IL-8 and TNF genes in bronchial biopsies before and after inhalation of VIP in COPD *in vivo*. Quantitative analysis of gene expression of IL-8 (A) and TNF (B) in human lungs. Total RNA was isolated from lung biopsies collected from patients before and after 3 months VIP treatment and from control subjects; mRNA levels of 18s were used for normalization. Data are expressed as mean $\pm$ SD (n=6). P Less-than sign 0.01 in treated versus untreated patients.

The expression of the two genes in biopsies of lung tissue by immuno-histochemistry demonstrated a similar pattern. While before the inhalation of VIP the expression of the two genes was more pronounced in tissue sections of COPD patients, following treatment the signal declined (Figure 4).

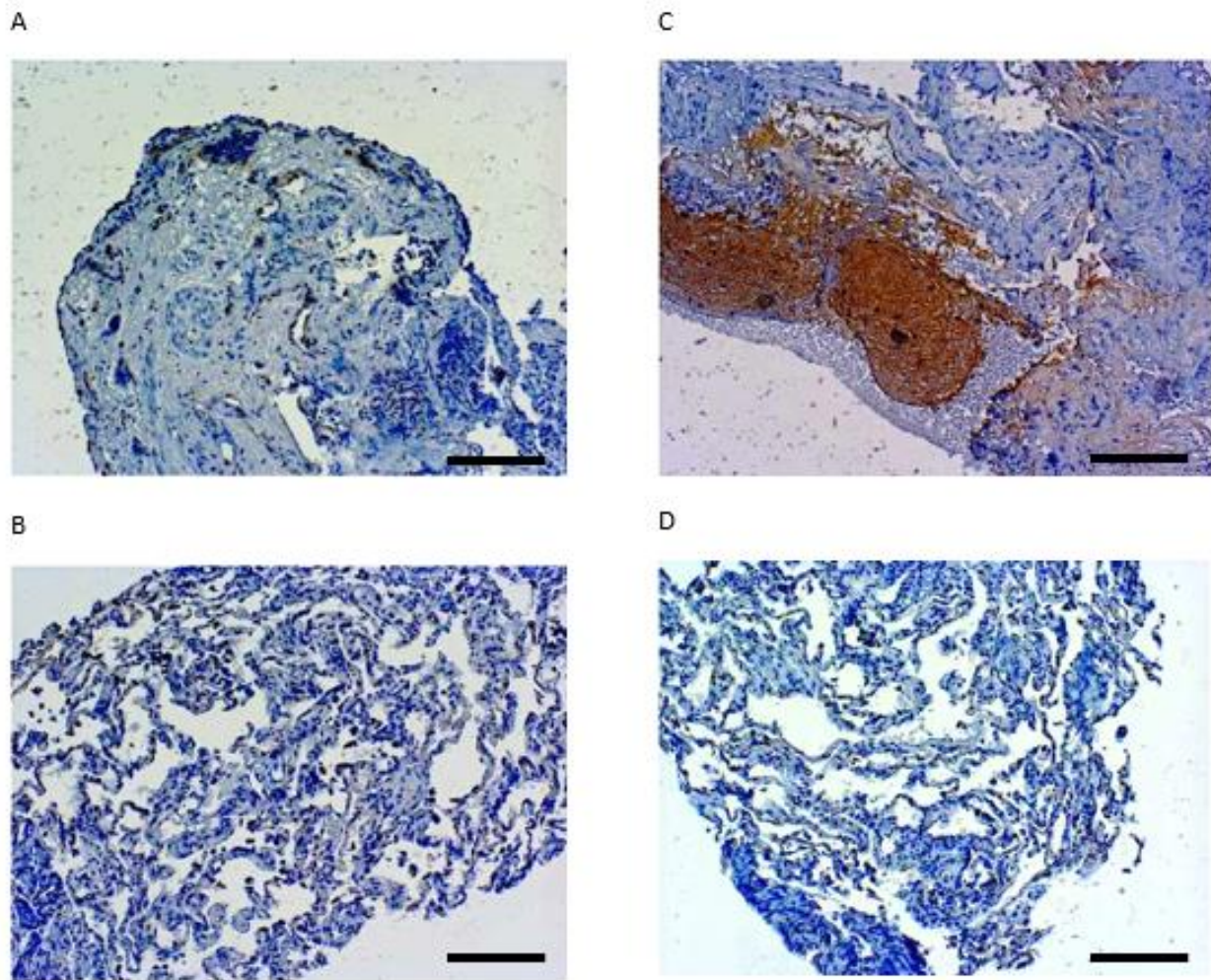
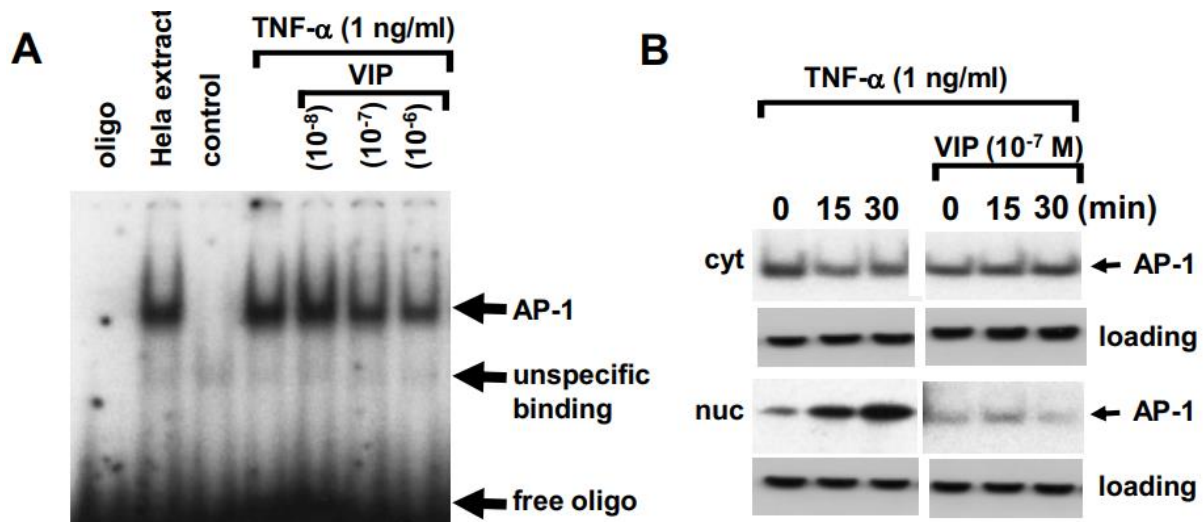


Figure 4: Expression of IL-8 and TNF in bronchial biopsies before and after inhalation of VIP in COPD. Immunohistochemical staining of IL-8 (A and B) and TNF (C and D) in lung tissue from COPD patients before (A and C) and after (B and D) 3 months VIP therapy. Bar ~ 200µm.

TNF activated the DNA binding of the three transcription factors AP-1, Stat3 and NF- κ B with individual kinetics as assessed by DNA mobility shift assay, and which was confirmed by the transfer of the respective proteins from the cytosol into the nucleus by immuno-blot analysis (Figure 5). When pre-incubated with VIP (10-6 M - 10-12 M) for 30 minutes, the DNA binding of AP-1 and Stat3 was significantly reduced in

a dose-dependent manner (Figure 5A, 5C) while that of NF- κ B was only marginally reduced (Figure 5E). A similar reducing effect of VIP was observed on the protein translocation of the respective transcription factors as shown for AP-1 in figure 5B, for Stat3 in figure 5D and for NF- κ B in figure 5F. Equal protein loading was confirmed by α -tubulin (loading).



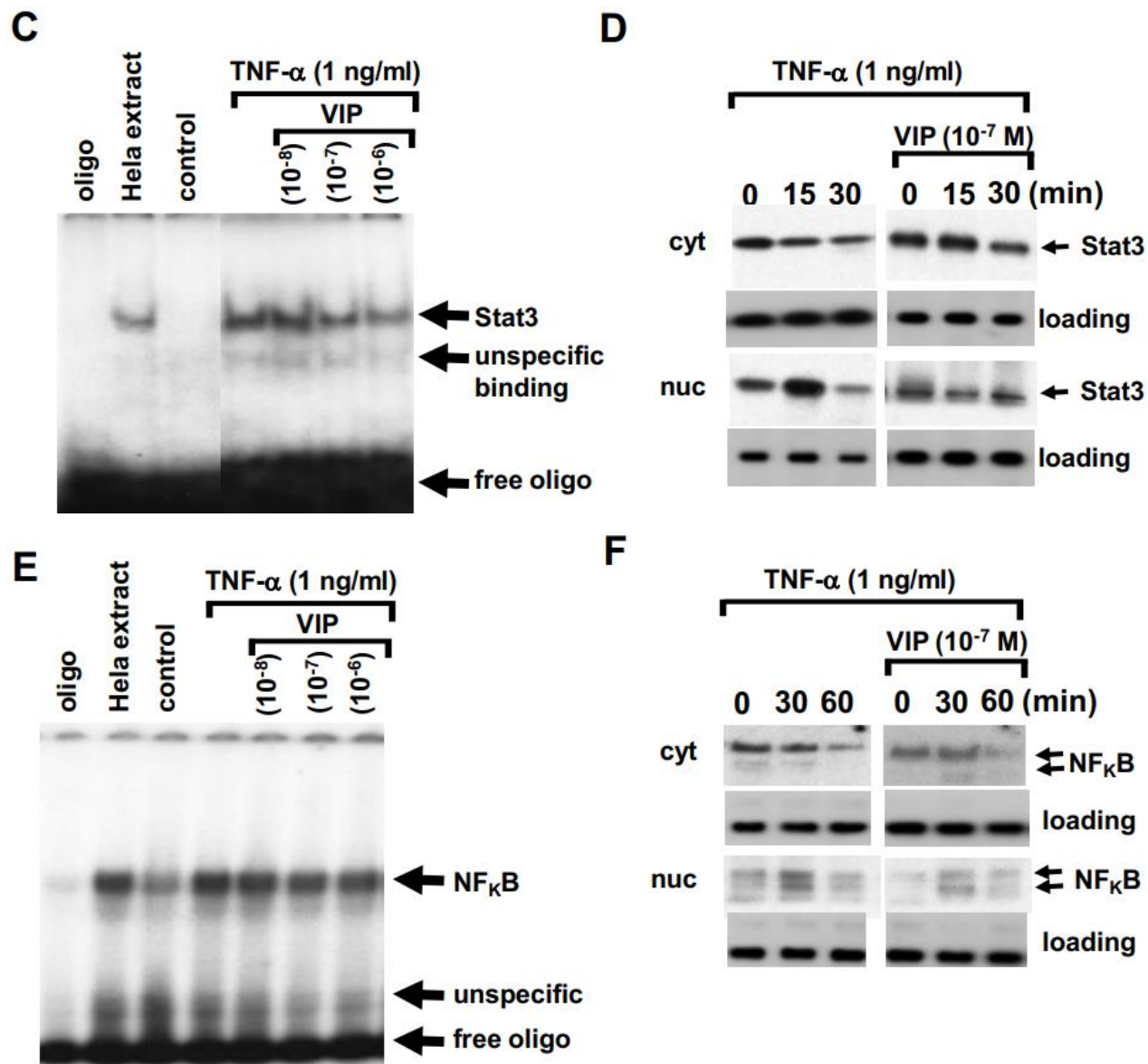


Figure 5: The effect of VIP on TNF induced transcription factor activity in primary BSMC and fibroblasts. A) A representative EMSA of the dose-dependent inhibition of TNF stimulated AP-1 to DNA binding by increasing concentrations of VIP. Similar results were obtained in two additional cell lines. B) A representative immuno-blot of the kinetic of TNF activated AP-1 translocation from the cytosol (cyt) into the nucleus (nuc) and its inhibition by VIP. Similar results were obtained in a second cell line. C) a representative EMSA of the dose- dependent inhibition of TNF stimulated Stat3 to DNA binding by increasing concentrations of VIP. Similar results were obtained in two additional cell lines. D) A representative immuno- blot of the kinetic of TNF activated Stat3 nuclear translocation and its inhibition by VIP. Similar results were obtained in a second cell line. E) a representative EMSA of the effect of VIP on TNF stimulated NF- κ B to DNA binding. Similar results were obtained in two additional cell lines. F) A representative immuno-blot of the effect of VIP on TNF activated NF- κ B nuclear translocation. Similar results were obtained in a second cell line.

A total of 34 participants were randomized out of 85 candidates who completed initial out-patient screening. The most common reason for exclusion was noncompliance and refusal of invasive diagnostic procedures. During the study, 4 subjects were withdrawn because of noncompliance with the study procedures: 2 in the VIP and 2 in the placebo group (Figure 6). Baseline characteristics of all 34 patients are provided in table 1. All COPD patients had moderate to severe reduced

pulmonary function with FEV1 before treatment of 1.33 ± 0.70 l in the placebo group and 1.42 ± 0.67 l in the VIP group, and the baseline lung function was comparable between both groups. More than 60% of the participants reported daily cough, more than 15% of the participants were smokers with a daily consumption of over 15 cigarettes, and 100% were former or current smokers.

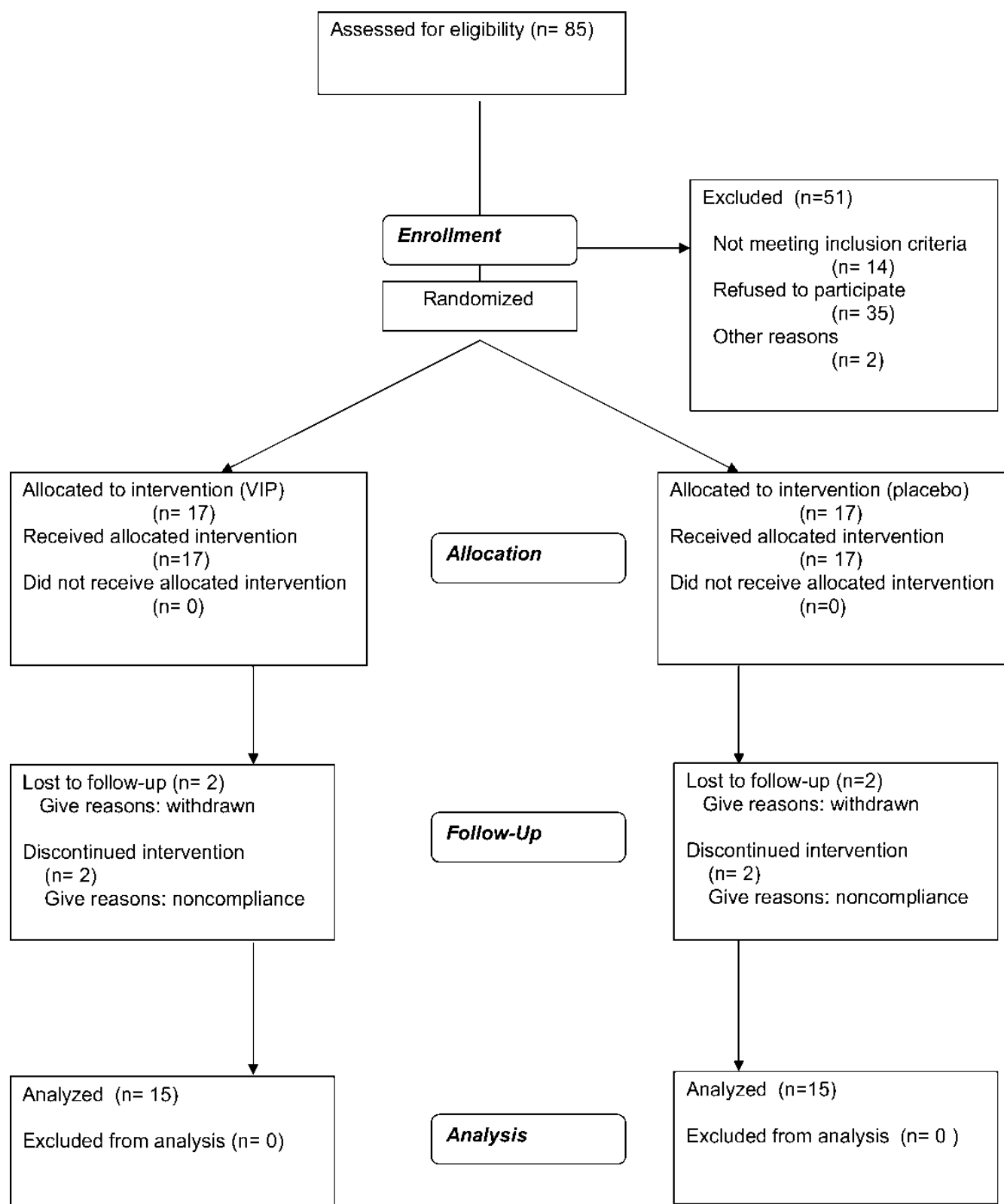


Figure 6: showing the flow of participants through each stage of the randomized trial.

The changes in post-bronchodilator FEV1 over time differed significantly (P Less-than sign 0,01) between the placebo and the VIP treatment group (Figure 7A and 7B). The placebo group showed a decline in FEV1 with a slope of -0.03 l (from 1.36±0.86 to 1.34±0.85) after 3 months, while VIP increased the FEV1 by 0.1 l (from 1.50 Plus-minus sign 0.68 to 1.60

Plus-minus sign 0.76) from baseline (P Less-than sign 0,01). Thus, the overall effect of VIP accounts for 0.13 l as compared to placebo (P Less-than sign 0.01). The post bronchodilator IVC increased by 0.155 l in the VIP group (Figure 7C) vs. baseline (P Less-than sign 0.05) and by 0.041 l in the placebo group (Figure 7D).

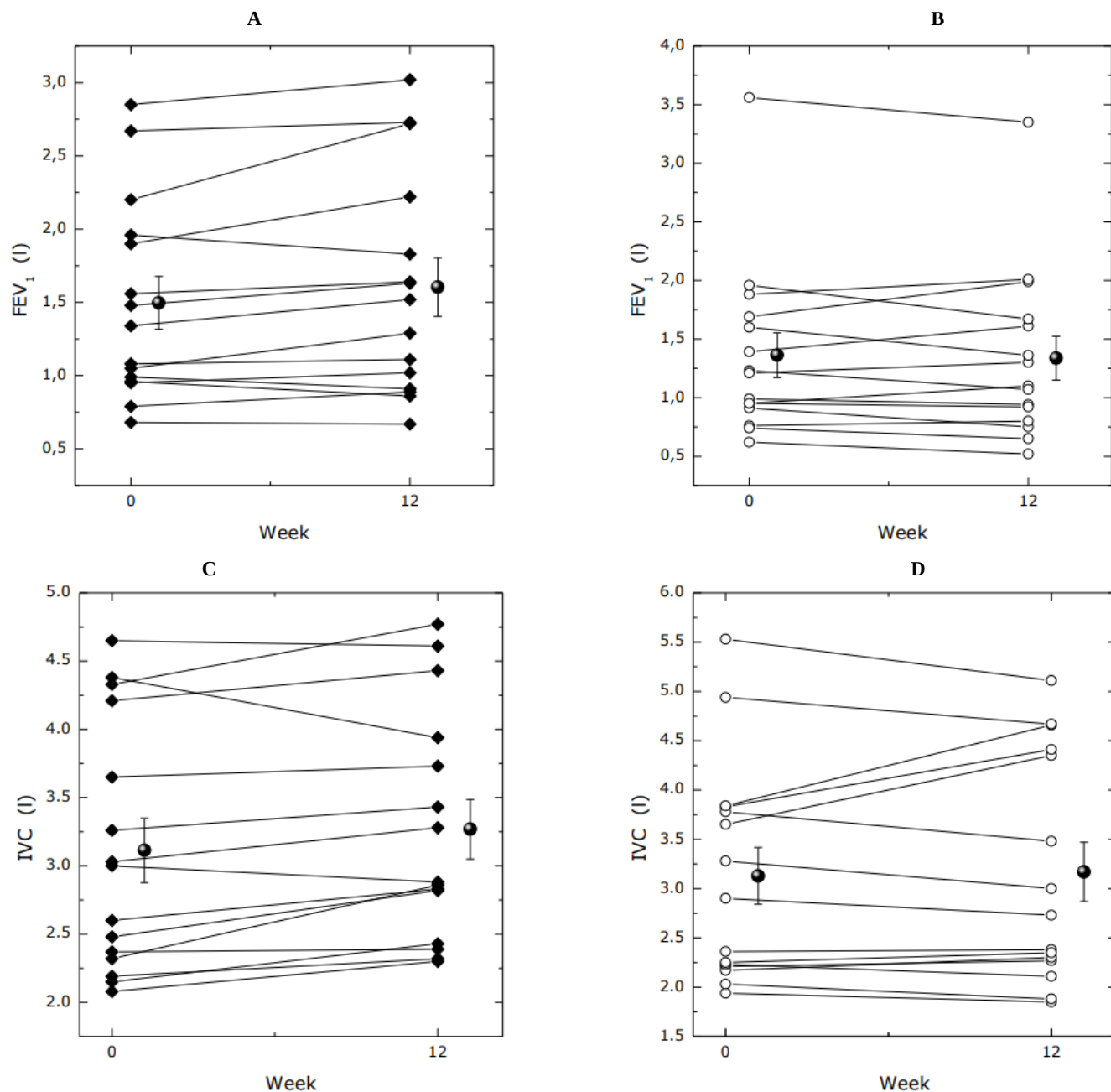


Figure 7: Forced Expiratory Volume in one Second (FEV1) $\pm$ SEM at selected time points (before and after 3 months treatment) by VIP (■) in group (A) and placebo (o) in group (B) (P Less-than sign 0,01). The numbers at each time point refer to data derived from VIP (■) or placebo (o) treatment group. Median change in Inspiratory Volume Capacity after 3 months treatment as compared to the value at randomization (Day 0) in the placebo (o) (D) or VIP group (■) (C).

As shown in table 1 (line 25), neither group of study patients had pulmonary hypertension. There was no significant change in pulmonary hemodynamic after treatment with VIP. (Data not shown).

Pos.	Parameter	Placebo Group (N=17)	VIP Group (N=17)	Difference ¹
1	Age (yr)	60.8 $\pm$ 6,7	57.4 $\pm$ 4,6	n.s.
2	Male sex no.(%)	10 (65%)	8 (48%)	n.s.
3	White race no.(%)	17 (100%)	17 (100%)	n.s.
4	PrebronchodilatorFEV1(litres)	1,33 $\pm$ 0,70	1,42 $\pm$ 0,67	n.s.
5	PrebronchodilatorFEV1(%predicted)	43,1 $\pm$ 19,20	45,3 $\pm$ 17,80	n.s.
6	FEV1:IVC ratio	42,6 $\pm$ 12,10	47,1 $\pm$ 13,30	n.s.
7	Cough - no.(%)	15(88,2%)	11 (64,7%)	n.s.
8	Sputum production - no.(%)	12 (71%)	11 (64,7%)	n.s.

9	Wheezing - no.(%)	8 (47,1%)	8 (47,1%)	n.s.
10	Smoked in past 3mo - no.(%)	6 (35%)	5 (29%)	n.s.
11	Total cigarettes smoking pack-yr	58,0± 39,6	52,4±37,8	n.s.
12	Regular medications - no.(%)	16 (94%)	16 (94%)	n.s.
13	Inhaled beta-adrenergic agonists - no. (%)	16 (94%)	16 (94%)	n.s.
14	Inhaled anti-cholinergic drug - no. (%)	16 (94%)	16 (94%)	n.s.
15	Oral beta-adrenergic agonists - no. (%)	0 (0%)	0 (0%)	n.s.
16	Theophylline - no.(%)	13 (76%)	10 (59%)	n.s.
17	Inhaled glucocorticosteroids - no.(%)	14 (82%)	15 (88%)	n.s.
18	Use of oxygen at home - no. (%)	2 (12%)	4 (23%)	n.s.
19	Other illness - no.(%)	17 (100%)	17 (100%)	n.s.
20	Diabetes mellitus - no.(%)	0 (0%)	1 (6%)	n.s.
21	History of ulcer - no.(%)	1 (6%)	2 (12%)	n.s.
22	Hypertension - no.(%)	10 (59%)	8 (47%)	n.s.
23	Disabling heart disease - no.(%)	1 (6%)	1 (6%)	n.s.
24	Disabling arthritis - no.(%)	0 (0%)	0 (0%)	n.s.
25	Mean pulmonary artery pressure (mmHg)	18.8±6,7	16.9±4,6	n.s.
26	Cardiac output (L/min)	5,4±0,6	5,7±1,3	n.s.
27	Pulmonary capillary wedge pressure (mmHg)	9,1±3,9	7,2±2,9	n.s.
29	Time from presentation to randomization days	20.6±4,71	22.6±4,71	n.s.

* Data are either “means ± SD” or “counts and (percent)”.

t Significance=P Less-than sign 0.05 for differences among groups by statistical analysis

(Students T-Test for continuous variables and by the chi-square test for categorical variables)

Table 1: Base-line characteristics of the patients

The six minutes walking distance increased by 24.1 m from 443.1 ± 90.8 to 467.2 ± 93.8 m after 12 weeks VIP treatment (P Less-than sign 0.05), and insignificantly decreased by 8.6 meters from 444.5 ± 115.5 to 436.0 ± 145.2 m after placebo (Figure 8A and 8B).

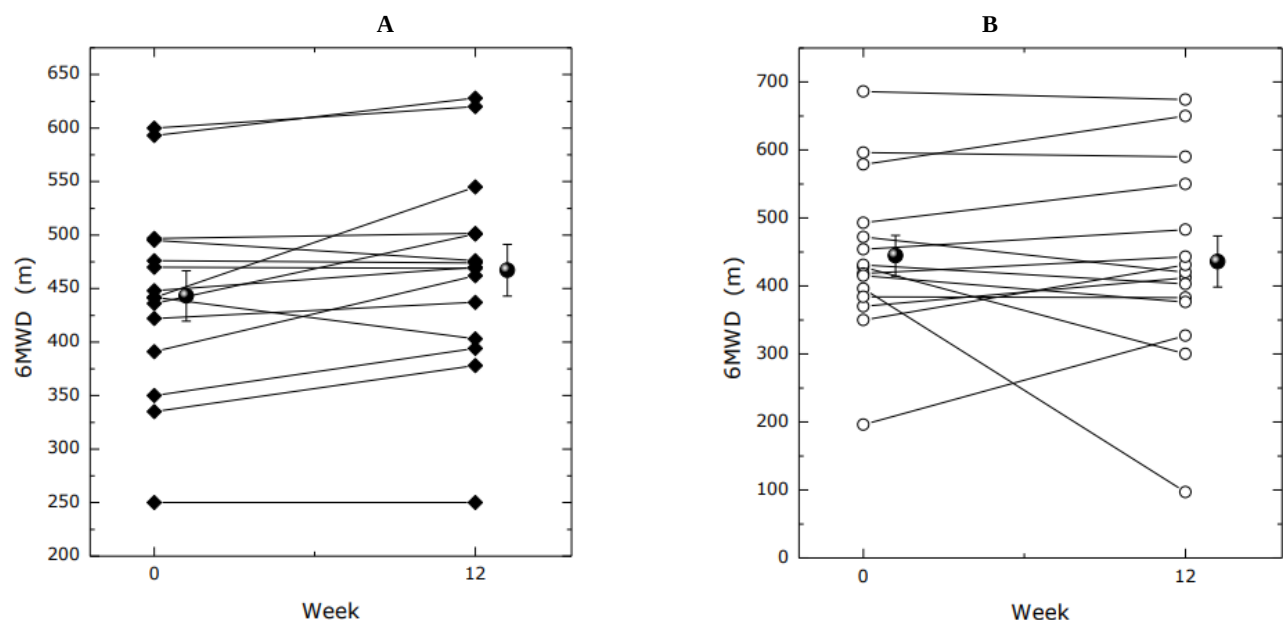


Figure 8: Change in Six-Minutes Walking Distance from baseline to day 90 in the VIP (■, A) group, and Placebo (○, B) group. P Less-than sign 0,05 for the comparison between VIP and placebo.

Borg dyspnea index, a primary variable, showed improvements (i.e., decreased score) with VIP (Figure 9). The changes from base-line in Borg dyspnea scale before 6MWT were -0,2 U for VIP (A), and +0,2 U for placebo (P Less-than sign 0,05) and -1,1 U for VIP, and 0,7 U for placebo after 6MWT (P Less-than sign 0,05).

Health-related quality of life assessed by SF-36, a primary variable (i.e., decreased score) and SGRQ showed improvements with VIP (Table 2). The changes from baseline in SF-36 - PHS were 4.08 for VIP and 0.20 for placebo (P Less-than sign 0.05). No significant changes were found in the mental health dimension in either group. In SGRQ the changes from baseline were -5.25 (from 46.52±4.06 to 41.27±4.35) for VIP and 1.72 for placebo (from 46.95±4.03 to 47.23±4.70, P Less-than sign 0.05).

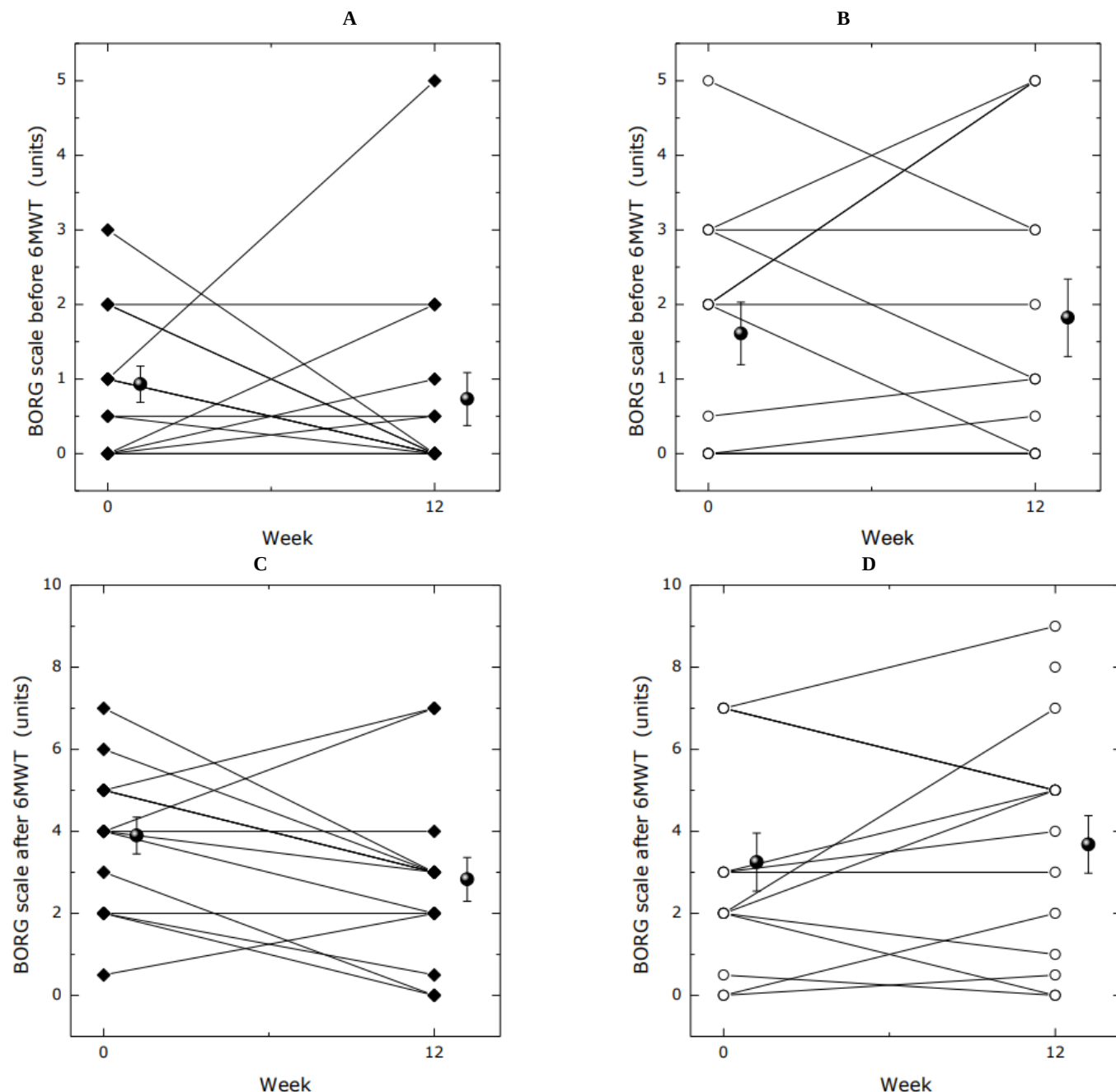


Figure 9: BORG dyspnea Score before (A and B) and after (C and D) 6MWT -exercise. Total scores from baseline to day 90 for the VIP (■) and placebo (○) groups. P Less-than sign 0,05 for comparison between VIP and placebo.

SF - 36 and SGRQ	VIP	Placebo
PHS at baseline	41.15±2.26	38.62±2.12
PHS after treatment (week 12)	45.23±2.07 (p=0,05)	38.82±2.49 (p=0,898)
MHS at baseline	44.45±2.41	44.89±2.53
MHS after treatment (week 12)	47.92±2.18 (p=0,366)	45.64±3.18 (p=0,796)
SGRQ at baseline	46.52±4.06	46.95±4.03
SGRQ after treatment (week 12)	41.27±4.35	47.23±4.70

* Data are either “means ± SD” or “counts and (percent)”.

t Significance=P Less-than sign 0.05 for differences among groups by statistical analysis (Students T-Test for continuous variables and by the chi-square test for categorical variables)

Table 2: Quality of life according to the short 36-item questionnaire - SF-36 and St. Georges respiratory questionnaire - SGRQ.

4. Discussion

VIP is released through inflammatory processes as evidenced by the finding that the serum VIP levels were significantly higher in COPD patients with acute exacerbations compared to stable COPD [27].

We found an inverse relationship between the serum concentration of VIP and the progression of COPD. VIP inhibited the release of IL-8 and TNF in parallel with the inhibition of transcription and expression of the two genes and downregulated the transcription factors AP-1 Stat3 and NF- κ B. The inhalation of VIP resulted in a significant improvement of lung function parameters of COPD patients (GOLD I-IV), a better quality of life and exercise capacity and in lesser exacerbations without relevant side effects. How can the efficacy of VIP in COPD be interpreted?

Firstly, an inverse relationship between the level of expression of VIP receptor proteins and of the level of circulating VIP was previously observed in pulmonary arterial hypertension [37]. Similarly, VIP serum levels in COPD patients were significantly decreased versus normal subjects suggesting a lack of VIP contributing to the pathogenesis of COPD. Elevated levels of several pro-inflammatory cytokines and other mediators have been demonstrated in the lungs of patients with COPD [8,17]. For example, IL-8, adhesion molecule (ICAM-1), and TNF are present in induced sputum [22], and their levels increased further during exacerbation [51]. The levels of TNF, IL-8 and metalloproteinase-9 (MMP-9) are elevated in bronchoalveolar lavage fluid from chronic smokers compared to non-smokers [24]. Blood levels of TNF, as well as of the soluble receptors for these cytokines, are two to three times higher in subjects with COPD than in the non-COPD population. Importantly, these levels correlate directly with the severity of airflow impairment [47, 13]. A recently performed study on gene expression profiles using serial analysis of gene expression and microarray analysis in lung biopsies of GOLD II smokers revealed a significantly increased expression of a number of genes encoding transcription factors or related proteins [31, 32]. Analogously, measurements of IL-8 and TNF mRNAs in bronchial biopsies of the COPD patients of our trial using real-time PCR demonstrated lower transcription after VIP treatment. Among its various immuno-modulatory properties, VIP was reported to down-regulate the biosynthesis of CXC- and CC-chemokines such as IL-8, ICAM-1, MMP-9, and TNF [11, 49]. Moreover, VIP inhibits the binding and activation of the transcription factor NF- κ B to corresponding promoter sequences of genes encoding mediators of inflammation, such as early growth factors 2/3 [11]. Another transcription factor which has been implicated in COPD is Stat3 [15, 38] and the observation that VIP inhibits TNF induced Stat3 activation in cells, as shown in this study, isolated from COPD patients indicates that VIP has a broad anti-inflammatory action. In mouse as well as in human fibroblasts cigarette smoke, the prevalent cause of COPD, activated AP-1 which was involved in the regulation of redox system-dependent pro-inflammatory factors such as heme oxygenase [2, 24]. Furthermore, it was suggested that the inhibition of NF- κ B and AP-1 could circumvent steroid insensitivity in chronic inflammatory diseases [1]. Since steroids are not very efficient in the therapy of COPD the inhibition of NF- κ B and AP-1 would explain the significant clinical improvement in COPD which we report here.

The molecular basis for the observed reduction of acute exacerbations following VIP treatment is unclear. Although various immuno-modulatory properties may be involved, experimental evidence suggests that VIP promotes Th-2 type and reduces inflammatory Th-1 type responses [10] that may lead to reduced susceptibility against infections.

Secondly, the fact that none of the patients included in this study exhibited pulmonary hypertension excludes the known vasodilatory effect of VIP as relevant effective mechanism. VIP has been shown to stimulate the dilatation of bronchial rings in vitro (Greenberg et al. 1987). Moreover, VIP inhalation resulted in increased FEV1 in asthmatics [33]. Similarly, in our trial the daily dose of 200 pg VIP significantly increased post-bronchodilator FEV1 and that of IVC throughout the 12-week treatment period ($P < 0.01$). This effect was achieved in addition to concomitantly applied β_2 -sympathomimetics, anti-cholinergics and inhaled corticosteroids. Studies using inhaled fluticasone propionate (500 ug twice daily), in a similar patient population - but without concomitant basic therapy - resulted in a 50 mL improvement in post-bronchodilator FEV1 over 6 months [33]. The combination of fluticasone plus salmeterol led to 65 mL or tiotropium bromide to 70 mL increase in FEV1 [7]. Since FEV1 is thought to predict prognosis and overall mortality in patients with COPD, the improvements in lung function recorded in our study are encouraging. Consistent with the results seen for post-bronchodilator FEV1, post-bronchodilator IVC increased with VIP which may explain - in part - the improvement of health-related quality of life. Thus, the SF-36 scores achieved with inhaled VIP compared to inhaled placebo, the peptide inhalation led to a significant improvement ($P < 0.05$).

Thirdly, chronic inflammation is generally regarded as a central mechanism in the pathogenesis of COPD [17]. Analogously, VIP has been shown to inhibit the inflammatory processes associated with COPD, and bronchial biopsy specimens of COPD patients (GOLD 0) exhibited increased expression of VPAC1 and VPAC2 receptors [28]. Furthermore, mitochondrial dysfunction and the effects of severely reduced availability of the protein deacetylase sirtuin-1 (SIRT1) are evident in COPD patients. The reduced SIRT1 expression within cells is a result of severe oxidative stress. The effects of VIP in pneumoplegic solution were tested in isolated rat lungs. Rat lungs were isolated, flushed and stored for 24 hours in presence or absence of VIP in the restoration solution. Electron microscopy showed that lungs stored in VIP-containing solutions had significantly more normal shaped mitochondria, less mitochondrial edema, less distortion of mitochondrial cristae, thinner basal lamina, and less aggregation of nuclear chromatin than control solutions.

Treatment with VIP also reduced histopathological severity of colitis and cell death markers in a murine model, leading to partial recovery of inhibited mitochondrial respiratory complexes, altered mitochondrial membrane potential and lowered ATP generation.

Mice lacking VIP (VIP-KO mice) gene develop right ventricular hypertrophy, thickened pulmonary artery, perivascular inflammatory cell infiltrates in the lung, and exhibit airway hyperresponsiveness to the cholinergic agonist methacholine. Treatment of these mice with exogenous VIP attenuates both, the vascular remodeling and right ventricular remodeling. [55, 56, 57, 58, 61, 62, 63, 64].

The fact that the treatment of COPD is limited to only few therapeutic options indicates the need for research and product development of additional alternatives. The unique ability of VIP to improve cardiopulmonary circulation and lung function in COPD, and quality of life, and 6 MWD without relevant side effects suggests that it may become a new therapeutic option. Future studies on VIP and VIP analogues are therefore necessary to reveal its full potential for clinical use.

Conclusion

Our study is the first to report that the inhalation of VIP has clear immunoregulatory effects in COPD patients. VIP inhalation might be an attractive, new immunoregulatory treatment strategy without systemic immunosuppression and negative side effects. Inhaled VIP can act alone, or in combination with any other approved COPD treatment option, with excellent safety profile. Further multicentric controlled studies are needed to evaluate the clinical efficiency of this new treatment option in COPD.

Author's Contributions

Wilhelm Mosgoeller, preclinical project design, histology, immunohistochemistry, Ventsislav Petkov, clinical study conceptualization, principal clinical investigator Bernhard Burian, patient recruitment and investigation, clinical study rollout Michael Roth, in vitro molecular laboratory studies, immunoblotting Christopher Lambers, in vitro cell isolation, sample administration Lutz-Henning Block, data review, manuscript writing, senior PI, Claudia Vonbank, clinical patient investigation and data administration

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Registration number for the clinical study:

ClinicalTrials.gov Identifier: NCT00464932

Scientific Knowledge on the Subject:

Molecular and clinical effects of the neuropeptide VIP in COPD.

What This Study Adds to the Field:

A new aspect of the role of Vasoactive Intestinal Peptide (VIP) in Chronic Obstructive Pulmonary Disease (COPD) and its therapeutical potential, when inhaled.

6. References

1. Adcock IM, Chou PC, Durham A, Ford P., (2009). Overcoming steroid unresponsiveness in airways disease" *Biochem Soc Trans* 37:(Pt 4)824-829.
2. Bagloli CJ, Sime PJ, Phipps RP., (2008). Cigarette smoke-induced expression of heme oxygenase-1 in human lung fibroblasts is regulated by intracellular glutathione. *Am J Physiol Lung Cell Mol Physiol* 295:(4) L624-636.
3. Bekdas M, Saygi B, Kilinc YB, Kilinc E., (2024). Plasma levels of neurogenic inflammation related neuropeptides in pediatric patients with community-acquired pneumonia and their potential diagnostic value in distinguishing viral and bacterial pneumonia" *Eur J Pediatr* 183:(4)1619-1627.
4. Burian B, Storka A, Marzluf BA, Yen YC, Lambers C, Robibaro B, Petkov V., (2010). Vasoactive intestinal peptide (VIP) receptor expression in monocyte-derived macrophages from COPD patients. *Peptides* 31:(4)603-608.
5. Busto R, Prieto JC, Bodega G, Zapatero J, Carrero I., (2000). Immunohistochemical localization and distribution of VIP/PACAP receptors in human lung" *Peptides* 21:(2)265-269.
6. Cai J, Chen Q, Mehrabi Nasab E, Athari SS., (2022). Immunomodulatory effect of N-acetyl seryl-aspartyl-proline and vasoactive intestinal peptide on chronic obstructive

- pulmonary disease pathophysiology. *Fundam Clin Pharmacol* 36:(6)1005-1010.
7. Cazzola M, Andò F, Santus P, Ruggeri P, Di Marco F, Sanduzzi A, D'Amato M., (2007). A pilot study to assess the effects of combining fluticasone propionate/salmeterol and tiotropium on the airflow obstruction of patients with severe-to-very severe COPD. *Pulm Pharmacol Ther* 20:(5)556-561.
8. Chung KF., (2001). Cytokines in chronic obstructive pulmonary disease. *Eur Respir J Suppl* 34:50s-59s.
9. Delgado M, Abad C, Martinez C, Leceta J, Gomariz RP., (2001). Vasoactive intestinal peptide prevents experimental arthritis by downregulating both autoimmune and inflammatory components of the disease. *Nat Med* 7:(5)563-568.
10. Delgado M, Ganea D., (2001). Inhibition of endotoxin-induced macrophage chemokine production by VIP and PACAP in vitro and in vivo" *Arch Physiol Biochem* 109:(4)377-382.
11. Delgado M, Ganea D., (2001). Inhibition of endotoxin-induced macrophage chemokine production by vasoactive intestinal peptide and pituitary adenylate cyclase-activating polypeptide in vitro and in vivo. *J Immunol* 167:(2)966-975.
12. Delgado M, Pozo D, Ganea D., (2004). The significance of vasoactive intestinal peptide in immunomodulation. *Pharmacol Rev* 56:(2)249-290.
13. Eid AA, Ionescu AA, Nixon LS, Lewis-Jenkins V, Matthews SB, Griffiths TL, Shale DJ., (2001). Inflammatory response and body composition in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 164:(8 Pt 1)1414-1418.
14. Fletcher C, Peto R., (1977). The natural history of chronic airflow obstruction" *Br Med J* 1:(6077)1645-1648.
15. Gao H, Ward PA., (2007). STAT3 and suppressor of cytokine signaling 3: potential targets in lung inflammatory responses. *Expert Opin Ther Targets* 11:(7)869-880.
16. Greenberg B, Rhoden K, Barnes PJ., (1987). Relaxant effects of vasoactive intestinal peptide and peptide histidine isoleucine in human and bovine pulmonary arteries. *Blood Vessels* 24:(1-2)45-50.
17. Groneberg DA, Chung KF (2004). Models of chronic obstructive pulmonary disease. *Respir Res* 5:(1)18.
18. Gunaydin S, Imai Y, Takanashi Y, Seo K, Hagino I, Chang D, Shinoka T (2002). The effects of vasoactive intestinal peptide on monocrotaline induced pulmonary hypertensive rabbits following cardiopulmonary bypass: a comparative study with isoproterenol and nitroglycerine. *Cardiovasc Surg* 10:(2)138-145.
19. Horstman DJ, McCall DA, Frank DU, Rich GF., (1999). Inhaled nitric oxide and nifedipine have similar effects on lung cGMP levels in rats. *Anesth Analg* 89:(4)932-937.
20. Johnson PR, Roth M, Tamm M, Hughes M, Ge Q, King G, Black JL., (2001). Airway smooth muscle cell proliferation is increased in asthma. *Am J Respir Crit Care Med* 164:(3)474-477.
21. Kapustnik VA, Istomina OV., (2019). The relationship of vip level with cardiopulmonary parameters in patients with chronic obstructive pulmonary disease in combination with hypertension. *Wiad Lek* 72:(7)1265-1268.
22. Keatings VM, Collins PD, Scott DM, Barnes PJ., (1996). Differences in interleukin-8 and tumor necrosis factor-alpha in induced sputum from patients with chronic obstructive

- pulmonary disease or asthma. *Am J Respir Crit Care Med* 153:(2)530-534.
23. Kostikas K, Papatheodorou G, Psathakis K, Panagou P, Loukides S., (2003). Oxidative stress in expired breath condensate of patients with COPD" *Chest* 124:(4)1373-1380.
 24. Li YT, He B, Wang YZ., (2009). Exposure to cigarette smoke upregulates AP-1 activity and induces TNF-alpha overexpression in mouse lungs. *Inhal Toxicol* 21:(7)641-647.
 25. Livak KJ, Schmittgen TD., (2001). Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods* 25:(4)402-408.
 26. Majo J, Ghezzi H, Cosio MG., (2001). Lymphocyte population and apoptosis in the lungs of smokers and their relation to emphysema. *Eur Respir J* 17:(5)946-953.
 27. Mandal J, Roth M, Costa L, Boeck L, Rakic J, Scherr A, Stolz D., (2015). Vasoactive Intestinal Peptide for Diagnosing Exacerbation in Chronic Obstructive Pulmonary Disease. *Respiration* 90:(5)357-368.
 28. Miotto D, Boschetto P, Bononi I, Zeni E, Cavallero G, Fabbri LM, Mapp CE., (2004). Vasoactive intestinal peptide receptors in the airways of smokers with chronic bronchitis. *Eur Respir J* 24:(6)958-963.
 29. Misaka S, Aoki Y, Karaki S, Kuwahara A, Mizumoto T, Onoue S, Yamada S., (2010). Inhalable powder formulation of a stabilized vasoactive intestinal peptide (VIP) derivative: anti-inflammatory effect in experimental asthmatic rats. *Peptides* 31:(1)72-78.
 30. Murray CJ, Lopez AD., (1997). Global mortality, disability, and the contribution of risk factors: Global Burden of Disease Study. *Lancet* 349:(9063)1436-1442.
 31. Ning W, Li CJ, Kaminski N, Feghali-Bostwick CA, Alber SM, Di YP, Choi AM., (2004). Comprehensive gene expression profiles reveal pathways related to the pathogenesis of chronic obstructive pulmonary disease. *Proc Natl Acad Sci U S A* 101:(41)14895-14900.
 32. Ning W, Lee J, Kaminski N, Feghali-Bostwick CA, Watkins SC, Pilewski JM, Choi AM., (2006). Comprehensive analysis of gene expression on GOLD-2 Versus GOLD-0 smokers reveals novel genes important in the pathogenesis of COPD. *Proc Am Thorac Soc* 3:(6)466.
 33. Paggiaro PL, Dahle R, Bakran I, Frith L, Hollingworth K, Efthimiou J., (1998). Multicentre randomised placebo-controlled trial of inhaled fluticasone propionate in patients with chronic obstructive pulmonary disease. International COPD Study Group. *Lancet* 351:(9105)773-780.
 34. Paredi P, Kharitonov SA, Barnes PJ., (2002). Analysis of expired air for oxidation products. *Am J Respir Crit Care Med* 166:(12 Pt 2) S31-37.
 35. Pauwels RA, Buist AS, Calverley PM, Jenkins CR, Hurd SS., (2001a). Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease. NHLBI/WHO Global Initiative for Chronic Obstructive Lung Disease (GOLD) Workshop summary. *Am J Respir Crit Care Med* 163:(5)1256-1276.
 36. Pauwels RA, Buist AS, Ma P, Jenkins CR, Hurd SS., (2001b). Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease: National Heart, Lung, and Blood Institute and World Health Organization Global Initiative for Chronic Obstructive Lung Disease (GOLD): executive summary. *Respir Care* 46:(8)798-825.
 37. Petkov V, Mosgoeller W, Ziesche R, Raderer M, Stiebelhner L, Vonbank K, Block LH., (2003). Vasoactive intestinal peptide as a new drug for treatment of primary pulmonary hypertension. *J Clin Invest* 111:(9)1339-1346.
 38. Qu P, Roberts J, Li Y, Albrecht M, Cummings OW, Eble JN, Yan C., (2009). Stat3 downstream genes serve as biomarkers in human lung carcinomas and chronic obstructive pulmonary disease. *Lung Cancer* 63:(3)341-347.
 39. Retamales I, Elliott WM, Meshi B, Coxson HO, Pare PD, Sciruba FC, Hogg JC., (2001). Amplification of inflammation in emphysema and its association with latent adenoviral infection. *Am J Respir Crit Care Med* 164:(3)469-473.
 40. Sandford AJ, Silverman EK., (2002). Chronic obstructive pulmonary disease. 1: Susceptibility factors for COPD the genotype-environment interaction. *Thorax* 57:(8)736-741.
 41. Seidel P, Merfort I, Hughes JM, Oliver BG, Tamm M, Roth M., (2009). Dimethylfumarate inhibits NF- κ B function at multiple levels to limit airway smooth muscle cell cytokine secretion. *Am J Physiol Lung Cell Mol Physiol* 297:(2) L326-339.
 42. Shapiro SD., (1999). The macrophage in chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 160:(5 Pt 2) S29-32.
 43. Shapiro SD., (2003). Proteolysis in the lung. *Eur Respir J Suppl* 44:30s-32s.
 44. Standardization of Spirometry, 1994 Update. American Thoracic Society. (1995) *Am J Respir Crit Care Med* 152:(3)1107-1136.
 45. Stockley RA., (2002). Neutrophils and the pathogenesis of COPD. *Chest* 121:(5 Suppl)151s-155s.
 46. Szema AM, Forsyth E, Ying B, Hamidi SA, Chen JJ, Hwang S, Gonzalez Bosc LV., (2017). NFATc3 and VIP in Idiopathic Pulmonary Fibrosis and Chronic Obstructive Pulmonary Disease. *PLoS One* 12:(1) e0170606.
 47. Takabatake N, Nakamura H, Abe S, Inoue S, Hino T, Saito H, Tomoike H., (2000). The relationship between chronic hypoxemia and activation of the tumor necrosis factor-alpha system in patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 161:(4 Pt 1)1179-1184.
 48. Tamm M, Roth M, Malouf M, Chhajed P, Johnson P, Black J, Glanville A., (2001). Primary fibroblast cell cultures from transbronchial biopsies of lung transplant recipients. *Transplantation* 71:(2)337-339.
 49. Tan YR, Qin XQ, Guan CX, Zhang CQ, Luo ZQ, Sun XH., (2003). Regulatory peptides modulate ICAM-1 gene expression and NF-kappaB activity in bronchial epithelial cells. *Sheng Li Xue Bao* 55:(2)121-127.
 50. Usdin TB, Bonner TI, Mezey E., (1994). Two receptors for vasoactive intestinal polypeptide with similar specificity and complementary distributions. *Endocrinology* 135:(6)2662-2680.
 51. Wedzicha JA, Seemungal TA, MacCallum PK, Paul EA, Donaldson GC, Bhowmik A, Meade TW., (2000). Acute exacerbations of chronic obstructive pulmonary disease are accompanied by elevations of plasma fibrinogen and serum IL-6 levels. *Thromb Haemost* 84:(2)210-215.

52. Wharton J, Davie N, Upton PD, Yacoub MH, Polak JM, Morrell NW., (2000). Prostacyclin analogues differentially inhibit growth of distal and proximal human pulmonary artery smooth muscle cells. *Circulation* 102:(25)3130-3136.
53. Wu D, Lee D, Sung YK., (2011). Prospect of vasoactive intestinal peptide therapy for COPD/PAH and asthma: a review. *Respir Res* 12:(1)45.
54. Zhong HL, Li PZ, Li D, Guan CX, Zhou Y., (2023). The role of vasoactive intestinal peptide in pulmonary diseases.
55. Martins IJ, Sirtuin, (2018). A diagnostic protein marker and its Relevance to Chronic Disease and Therapeutic Drug Interventions" *EC Pharmacology and Toxicology* 6.4: 209-215.
56. Martins IJ., (2017). Nutrition Therapy Regulates Caffeine Metabolism with Relevance to NAFLD and Induction of Type 3 Diabetes. *J Diabetes Metab Disord* 4: 019.
57. Martins IJ., (2017). Single Gene Inactivation with Implications to Diabetes and Multiple Organ Dysfunction Syndrome. *J Clin Epigenet.* 3:24.
58. Martins IJ., (2016). Anti-Aging Genes Improve Appetite Regulation and Reverse Cell Senescence and Apoptosis in Global Populations. *Advances in Aging Research*, 5, 9-26.
59. Leuchte H, Baezner C, Baumgartner RA, Bevec D, Bacher G, Neurohr C, Behr J., (2008). Inhalation of vasoactive intestinal peptide in pulmonary hypertension" *Eur Respir J.* 32: 1289-1294.
60. Prasse A, Zissel G, Lützen N, Schupp J, Schmiedlin R, Gonzalez-Rey E, Rensing-Ehl A, Bacher G, Cavalli V, Bevec D, Delgado M, Müller-Quernhei J., (2010). Inhaled Vasoactive Intestinal Peptide Exert Immunoregulatory Effects in Sarcoidosis. *Am J Respir Crit Care Med.* 182, 540-558.
61. Yanagisawa S, Papaioannou AI, Papaportfyriou A, Baker JR, Vuppusetty C, Loukides St, Barnes PJ, Ito K (2017). Decreased Serum Sirtuin-1 in COPD. *Chest* 152(2):343-352.
62. Allessandrini F, Thakkar M, Foda HD, Said SI, Lodi R, Pakbaz H, Schraufnagel DE., (1993). Vasoactive intestinal peptide enhances lung preservation. *Transplantation* 56, 964-973.
63. Spoorthi BC, More SS, Gautham SA, Ghosh S, Saha I, Maiti AK., (2020). Role of free radical scavenging activity of vasoactive intestinal peptide in the attenuation of mitochondrial dysfunction to ameliorate dextran sulphate sodium-induced colitis in mice: Implications in ulcerative colitis. *J Dig Dis.* 1-13.
64. Szema AM, Hamidi SA, Lyubsky S, Dickman KG, Mathew S Abdel-Razek T, Chen JJ, Waschek JA, Said S., (2006). Mice lacking the VIP gene show airway hyperresponsiveness and airway inflammation, partially reversible by VIP". *Am J Physiol Lung Cell Mol Physiol* 291: L880-L888



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