

## **Journal of Nutrition and Food Processing**

Zakari David Adeiza \*

Open Access Research Article

# Microbiological Quality of Selected Non-Sterile Pharmaceutical Products Retailed in Anyigba, Kogi State, Nigeria

Adejo Patience Omebije  $^1$ , Zakari David Adeiza  $^{1,3}$ , Sule Queen  $^1$ , Edegbo Emmanuel  $^1$ , Akoh Phoebe Queen  $^1$ , Olorunmowaju Abiodun Israel  $^1$ , Shuaib Yusuf Danjuma  $^2$ 

- <sup>1</sup> Department of Microbiology, Prince Abubakar Audu University, PMB 1008, Anyigba, Kogi state, Nigeria
- <sup>2</sup> Department of Science Laboratory Technology, Kogi State Polytechnic, PMB 101, Lokoja, Kogi State, Nigeria
- <sup>3</sup> Department of Microbiology, Federal University of Technology, PMB 2240, Abeokuta, Ogun State, Nigeria.
- \*Corresponding author: Zakari David Adeiza, 1Department of Microbiology, Prince Abubakar Audu University, PMB 1008, Anyigba, Kogi state, Nigeria.

Received date: June 27, 2023; Accepted date: November 03, 2023; Published date: November 10, 2023

**Citation:** Adejo P. Omebije, Zakari D. Adeiza, Sule Queen, Edegbo Emmanuel, Akoh P. Queen etc. (2023), Microbiological Quality of Selected Non-Sterile Pharmaceutical Products Retailed in Anyigba, Kogi State, Nigeria, *J. Nutrition and Food Processing*, 6(9); **DOI:10.31579/2637-8914/165** 

**Copyright:** © 2023, Zakari David Adeiza. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

#### **Abstract:**

This study was designed to check for the microbiological quality of non-sterile products retailed in Anyigba. The use of contaminated non-sterile pharmaceutical products can cause hazards in a majority of ways like economic loss to the industrialists, alter the therapeutic effect of the drug and affect the health of a patient. A total of 10 samples were collected; the isolated organisms were characterized and identified by using morphological, cultural and biochemical tests. The organisms isolated were Staphylococcus aureus, E.coli and Pseudomonas spp, Aspergillus spp, Mucorspp, Saccharomyces spp and Rhizopus spp. The percentage of the isolate includes S. aureus 37 (45.7%), E.coli 16 (19.7%) Pseudomonas spp 28 (34.5%) to safeguard the product from contamination, it is important to ensure that good manufacturing practices such as raw material testing, equipment sanitization and automation, microbial testing of water, training of personnel, post marking surveillance, monitoring of environment among others were employed. The focus should also be on surveillance and effective monitoring of the distribution and marketing of pharmaceutical products. Local regulatory agencies such as the Standard Organization of Nigeria (SON), and the National Agency for Food,drug administration and Control (NAFDAC) should always ensure that all pharmaceuticals released into the market for sales and consumption should conform to specifications and are fit for their intended use.

**Key words:** staphylococcus aureus; E.coli; pseudomonas spp; SON;NAFDAC

#### Introduction

Non-sterile drugs are pharmaceutical products that are not completely free from viable microorganisms or contamination due to the environment in which there are produced and the raw materials used in their formation. Examples of non-sterile drugs: solutions, ointments, creams, powders, capsules and tablets [17].

The pharmaceutical industry is an important element of health all over the world, [17]. Pharmaceutical products are divided into sterile and non-sterile products. Non-sterile pharmaceuticals are not produced by aseptic processes and therefore, are not expected to be free from the degree of contamination in non-sterile products is regulated and is based on the acceptance criteria for microbiological quality established in pharmacopeia monographs [17].

The poor qualities of medicines are not only a health hazard but also a waste of money for both government and consumers [4]. Therefore, the

maintenance of quality with continuous improvement in facilities is very important in pharmaceutical industries [4]. To achieve the quality objective, it is necessary to control all stages of drugs, which covers all matters, which individually or collectively influence the quality of a product, including raw materials, manufacturing the process and the evaluation of the finished product [15].

One of the control stages is the assessment of the microbiological quality of medicinal products [16]. Syrups are viscous oral liquids that may have one or more active ingredients in solution which usually contain large amounts of sucrose or other sugars to which certain polyhydric alcohols may be added to inhibit crystallization or to modify solubilization, taste and other properties [17]. Sugarless syrups may contain sweetening and thickening agents with 95% ethanol being a preservative solvent that incorporates agents, in addition, antimicrobial agents are also added to

syrups [17]. The presence of microbes in syrups is a great public health concern globally [17].

Contamination of pharmaceutical preparations with microorganisms irrespective of whether being pathogenic or non-pathogenic can bring about changes in the drugs' physical characteristics, including the breaking of emulsions, fermentation of syrups, and appearance of turbidity or deposit; besides producing possible odours and colour changes. The source of contamination may be from start-up materials, water used during manufacturing, operational equipment, the untidy surrounding environment and through workers, the pharmaceutical manufacturing and packaging environment, raw materials as well as the manufacturing water may attribute to the microbiological spoilage of the finished products [4,6,11]

The presence of a high number of non-pathogenic microorganisms in pharmaceutical products is objectionable as the organisms may deteriorate active ingredients and interfere with the desired activity of the product or generate toxic metabolites [6]. Since non-sterile pharmaceuticals are not produced by aseptic processes and, thus not expected to be free from microbial contaminations which can lead to significant economic loss to the industry as well as morbidity and mortality of the consumers [8].

## **Materials and Method**

#### **Inclusion Criteria**

This research strictly examines the quality of non-sterile pharmaceutical drugs in the form of tablets, and syrup with various routes of administration and compositions.

#### **Exclusion Criteria**

The microbiological quality of sterile drugs will not be inclusive

## **Sample Collection**

The samples to be used include 10 non-sterile drugs which include: Panadol extra, Nifedipine,Ampicillin, Vitamin C, Paracetamol, Diclofenac, Metronidazole,Emzolyn cough syrup,Tutolin Cough Syrup, Moduretic randomly selected from different pharmaceutical retails in Anyigba.

#### Isolation and quantification of microbial contaminants

Selective and nonselective culture media will be used for quantification and isolation of the microbial contaminants, which are nutrient agar, Saboraud dextrose agar, and MacConkey agar. A 1 mL aliquot of each suspension will be directly plated onto the sterile media and incubated for 24–48 hours at 370c [12]. Pure and single microbial colonies will be subcultured onto solid and liquid media, and incubated at 37°C for 24 hours, and the substantive isolates will be finally stored at 4°C until further use.

## **Identification of microbiological** contaminants

Isolated microbial contaminants will be subjected to standard microbiologic identification tests [5]. This includes;

For bacteria: Gram Staining, Indole test, Citrate utilization test, Catalase test, Coagulase test, Methyl- Red Test, Urease Test and motility test

For fungi: Isolated organisms will be subjected to standard microbiologic identification tests based on cultural characteristics and colony growth morphologies.

#### Result

Colony forming unit (cfu/ml/g) of the samples on Nutrient agar and MacConkey agar  $\,$ 

Table 1 shows the bacteria load of the samples analyzed on Nutrient agar and Macconkey agar. Sample A had the highest bacteria load of 1.8 x103cfu/ml/g and Sample H have the lowest bacteria load of 1 x102.

Sample	NA(cfu/ml/g)	MAC(cfu/ml/g)		
A	$1.8 \times 10^{3}$	-		
В	3 x 10 <sup>2</sup>	-		
С	$4.2 \times 10^{2}$	-		
D	$1.2 \times 10^{3}$	-		
E	$6.3 \times 10^{2}$	-		
F	-	-		
G	-	2 x 10 <sup>2</sup>		
Н	-	$1 \times 10^{2}$		
I	-	-		
J	-	-		

**Key:** (-) = no growth,

NA - Nutrient Agar, MAC - MacConkey Agar,

A – Panadol extra, B – Vitamin C, C – Nifedipine, D – Ampicillin,

E – Metronidazole, F – Diclofenac, G – Paracetamol, H – Moduretic, I – Emzolyn, J – Tutolin

Table 1: Colony forming unit (cfu/ml/g) of the samples on Nutrient agar and MacConkey agar

# Colonial Morphology, Gram Reaction and Biochemical Characteristics Of Bacteria Isolates Sample

Table 2 shows the colony morphology of the different isolates according to their shapes, colour, elevation, edge, consistency, and colony surface. The gram reaction shows the isolate's ability to retain the primary dye (crystal violet) which classifies them as gram-positive and gram-negative

organisms. Also, different biochemical test was used for the identification of the various isolates.

## Morphology

### **Biochemical test**

Isolat e	Shape	Colour	Elevatio n	Edge	Consistenc y	Colon y surfac e	Gram reactio n		In d	Mo t	Co a	Me t	U r	Ci t	Probable organisms
1	Circula r	Milky	Raised	Entire	Moist	Smoot h	+	+	-	+	+	+	+	+	S. aureus
2	Circula r	Yellow	Raised	Entire	Moist	Smoot h	-	-	+	+	-	+	-	-	E coli
3	Circula r	Milky	Raised	Entire	Moist	Smoot h	+	+	-	-	+	+	+	+	S. aureus
4	Circula r	Milky	Raised	Entire	Moist	Smoot h	+	+	-	-	+	+	+	+	S. aureus
5	Circula r	Greenis h	Flat	Irregula r	Moist	Smoot h	-	+	-	+	-	-	-	+	<i>Pseudomonas</i> sp p
6	*	*	*	*	*	*		*	*	*	*	*	*	*	
7	Circula r	Milky	Raised	Entire	Moist	Smoot h	+	+	-	-	+	+	+	+	S. aureus
8	Circula r	Milky	Raised	Entire	Moist	Smoot h	+	+	-	-	+	+	+	+	S. aureus
9	*	*	*	*	*	*	*	*	*	*	*	*	*	*	
10	*	*	*	*	*	*	*	*	*	*	*	*	*	*	

 $Key\ Cat = Catalase\ Met = Methylred(-) = positive,\ Ind = Indole\ Ur = Urease\ (-) = negative\ (*) = no\ growth,\ Mot = motility,\ Cit = Citrate\ ,\ Coa=Coagulase$ 

Table 2: Colonial Morphology, Gram Reaction and Biochemical Characteristics of Bacteria Isolates from Samples

## Microbial Counts of Non-Sterile Drugs Samples

Table 3 shows the accepted microbial limits of bacteria for non-sterile Pharmaceutical products [18] Sample A wasmicrobiologically unacceptable because the Total Viable Bacteria Count exceed the limits

103cfu/g/ml. Sample B was also microbiologically unaccepted because the specified organism E.coi is present, Sample D was microbiologically unacceptable because the Total Viable Bacteria (TVBC) exceed the accepted. While other samples were microbiologically acceptable.

Drug S	Sample	TVBC	Specified microorganisms ( E. coil)	Comments
	A	1.8 x 10 <sup>3</sup>	Absent	Microbiologically unacceptable
	В	$3 \times 10^{2}$	Present	Microbiologically unacceptable
	С	$4.2 \times 10^{2}$	Absent	Microbiologically Acceptable
	D	$1.2 \times 10^3$	Absent	Microbiologically unacceptable
-	E	6.3 x 10 <sup>2</sup>	Absent	Microbiologically Acceptable
F	G	$2 \times 10^2$	Absent	- Microbiologically Acceptable
	Н	$1 \times 10^{2}$	Absent	Microbiologically Acceptable
I		-	-	-
J		-	-	-

Table 3: Microbial Counts of Non-Sterile Drugs Samples

Accepted microbial limits of bacteria for non-sterile oral drugs [18]

KEY: TVBC = total viable bacteria count (-) = no growth

(\*102 cells/ml) = the Acceptable microbial limits of bacteria for syrup

(\*103 cells/g) = the Acceptable microbial limits of bacteria for the tablet.

Frequency Occurrence of Bacteria Isolate from Non-Sterile Pharmaceutical Products

Isolates	%	%	%	%	%	%	%	Total
	A	В	C	D	E	G	Н	
S.aureus	10 (58.8)	6 (42.9)	4 (40	3 (30)	0	6 (60)	8 (66.6)	37 (45.7%)
E. coli	2 (11.8)	5 (35.7)	2 (20)	3 (30)	1 (12.5)	1 (10)	2 (16.7)	16 (19.7%)
Pseudomonas	5 (29.4)	3 (21.4)	4 (40)	4 (40)	7 (87.5)	3 (30)	2 (16.7)	28 (34.5%)
spp.								

Key: A – Panadol extra, B – Vitamin C,C – Nifedipine, D– Ampicillin,

**Table 4:** shows the frequency of occurrence of bacterial isolates. Staphylococcus aureus had the highest frequency with 37(45.7%) and E.coli had the least frequency with 16(19.7%)

## Colonial Morphology of Fungi Isolated from Non-Sterile Pharmaceutical Products.

Table 5 shows the colonial morphology of fungi isolated from the nonsterile pharmaceutical product according to their spore, septate or nonseptate hyphae, texture, and description of the colony on media. The probable organisms isolated include Aspergillus spp, Mucor spp, Penicillium spp, Rhizopus spp and Saccharomyces spp.

Isolate	Description of colony	Septate/Aspetate	Spore	Texture	Probable organisms	
1	Greyish black spore with white margin	Septate	Sporulating	Powdery	Aspergillus spp	
2	-	-		-		
3	Whitish cream raised and smooth colony	Aseptate	Sporulating	Velvety	Mucorspp	
4	Green spores with white margin	Septate	Sporulating	Powdery	Penicilliumspp	
5	Whitish cotton –like with black spores	Aseptate unbranched	Round and black	Velvety	Rhizopus spp	
6	-	-	-	-	-	
7	Whitish cotton -like with black spores	Aseptateunbranched	Round and black	Velvety	Rhizopusspp	
8	Small cream colonies that are raised and smooth	Aseptate	Budding cells	Velvety	Saccharomyces spp	
9	-	-	-	-	-	
10	-	-	-	-	-	

Key:(-) = No Growth

 Table 5: Colonial Morphology of Fungi Isolated from Non-Sterile Pharmaceutical Product.

## **Discussion**

The result of this study showed that the microbial load in tablets is high while the syrup produces no growth. They include sample A (Panadol

extra) is 1.8x103, B (Vitamin C) is 3 x105, (Nifedipine) is 4.2 x102, D ampicillin 1.2 x103, E (metronidazole) 6.3 x102, G (Emozor para),2.0 x102, and H (moduretic) 1x102 from work sample A, B and D are

E- Metronidazole, F - Diclofenac, G - Paracetamol, H- Moduretic,

I – Emzolyn, J – Tutolin.

microbiologically unacceptable because the total aerobic microbial count is greater than the standard acceptable limit.

According to the acceptance criteria of non-sterile dosage from (European pharmacopoeia 2010). Total anaerobic microbial count (cfu/g/ml) is 103 and total yeast and mould count (cfu/g/ml) 102 specified that E. coli is present while samples F (diclofenac), I (cough syrup) and J( totulin cough syrup) have no contamination which may be due to proper hygienic practices, good manufacturing practices, proper storage and proper handling and packaging.

This work is similar to [3, 13]. All the samples examined the standard limit for non-sterile preparation which agrees with the work.

isolated from this work include Staphylococcus Aureus, E. coli And Pseudomonas while the fungi include Aspergillus spp, Mucorspp, Saccharomyces spp and Rhizopusspp. Staphylococcus aureus has the highest frequency of occurrence of 37(45.7%) This is the same as the finding of [14] the above list organism was also isolated, this work also agrees with the work of (Obueke, 2016) who observed that Staphylococcus aureus was the bulk of microbial contamination of the result of this survey show contamination of drug sample which could be from varying sources this finding agrees with the report of [10]. This could be a result of the water, environment, human resources, source from packaging as well as its natural surface flora, the skin and the respiratory tract could be the possible source of contamination, material and excipient and personnel [10]. These entire factors may account for the incident of drug contamination observed in this study.

[9] reported that Staphylococcus aureus and E. coli are the major contaminants of pharmaceutical products agree with this work.

Fungi isolated were Aspergillus spp, Mucorspp, Penicillium spp, Rhizopussppand Saccharomyces spp [1] and [7] carried out quality testing on non-sterile drugs and he also isolated E.coli, Staphylococcus aureus which was among the organism isolated in this work.

The fungi isolated are Aspergillus niger, Mucorspp, Saccharamycesspp and Rhizopusspp. Where the same as fungi isolated in the findings of [2] also correlate with the finding of [15].

#### Conclusion

Most of the brand's non-sterile pharmaceutical products sold in outlets in Anyigba were found to contain various levels of microbial contaminants which may constitute a public health concern and economic problem. A significant number of microorganisms isolated from the sample were either from human sources or from airborne. Local regulatory agencies such as the Standathe rd Organization of Nigeria (SON), and the National Agency for Food, drug administration and Control (NAFDAC) should always ensure that all pharmaceuticals products released to the market for consumption and sales should conform to specifications and fit for their intended use.

Non-sterile preparations have less stringent requirements regarding the exclusion of microbes. They need not be sterile but it has to be shown that some specifically named organisms are not present in them [18].

#### References

- Adetunji, M., Kilani, K. and Olaifa, W. (2017). Microbiological Quality of Selected Non-sterile Pharmaceutical Products. Microbiology Journal. Volume 3(1):339-346
- 2. Aha, R., Alai, E. and Sara, A. (2018). Microbiology quality control of some non-sterile preparations commonly used in

- Pakistan. Journal of Pharmaceutical Science. Volume 31:1237-1242.
- Akerele, O.J., Ukoh, G.C. (2002) Aspects of microbial contamination of tablets dispensed in hospitals and community pharmacies in Benin City, Nigeria. The pharmaceutical and chemical Journal. Volume 2:224-230
- Bhaskar, M., Bhattacharya, S. and Yadav, A. (2011). Total Quality Management inpharmaceuticals. International Journal of Pharm Tech Research. Volume 3(1):365 –37
- Clinical Laboratories Standards Institute.2010Performance Standardsfor Antimicrobial Susceptibility Testing; Sixteenth Informational Supplement. Wayne, PA: Clinical Laboratories Standards Institute; pdf. Accessed Jul 20:24-465.
- Gad, G.F.M., Aly, R.A.I. and Ashour, M.S.E. (2011). Microbial evaluation of some non-sterile pharmaceutical preparations commonly used in the Egyptian market. Tropical Journal of Pharmaceutical Research. Volume 10 (4): 437-445.
- Gamal-Fadi., M., Gad-Reham, A., Ibrahem, A. and Muhammed, S.E. (2010). Microbiology quality control of some non-sterile preparations. Tropical Journal of Pharmaceutical research. Volume 10(4):437-445.
- Hanlon, G. and Hodges, N. (2013). Essential Microbiology For Pharmacy and Pharmaceutical Science. First edition, School of Pharmacy and Biomolecular Sciences, University of Brighton, UK. A John Willey & Sons, Ltd., Publication, UK. Volume 137:163 – 167
- 9. Hossain, M., Shamim, A and Rahman, M.Z. (2004). Quantitative examination of aerobic bacteria and fungi in locally available antacid suspension and possible contamination by specified bacteria. Pak. J. Bio. Sci. 7(11): 2014-2017.
- 10. Hugo, W.B. and Russell, A.D. (1983). Pharmaceutical microbiology; 3rd edition,pp.340-343,354-361.
- Kabir, M. S. and Dulal, M. H. 2013. Microbiological Quality Assessment of Vitamin B syrups and Antibiotics Susceptibility Profile of the Isolated Esherichiacoli. IOSR Journal of Pharmacy and Biological Sciences. Volume 8(4): 61 – 64
- Konemar, E.W., Allen, S.D., Janda, W.M., Schreekenberger, P.C. and Winn, W.C. (1992). The Enterobacteriaceae. In: Color Atlas and Text Book of Diagnostic Microbiology.
- Mwambete, K.D. Justin-Temu, M. and Fazleabbas, S.f. (2009).
   Microbiological assessment of commercially available quinine syrups and water for injection in Dar es Salaam, Tanzania. Trop.
   J. Pharm. Res. Obuekwe CO. obuekwe IF, Rafiq M. (2000). Surface contamination in some pharmaceutical products. Journal of Applied Microbiology. Volume 3:34-57
- 14. O-Day H. and kamil, D. L. (2011) farmaciav.Volume 59(2):133-146
- Ratajcza, M., Kubicka, M.M., Kaminska, D., Sawicka, P. and Dludaszewka, J. (2014) Microbiological quality of non-sterile pharmaceutical products. Saudi pharmaceutical Journal. Volume 23:303 – 307
- Tyski, S. 2011. How to manufacture pure and safe drug.Pharmaceutical Industry.46(4):57-61
- 17. Uddin, M., Mamun, A., Akter, N., Sarwar, S., Rashid, M. and Amran, S. (2016). Pharmaceutical Industry. 46:68-478.
- United States Pharmacopeial Convention. (2013). Microbial Limits Tests- Nutritional Supplements. In U.S Pharmacopeia Rockville, Maryland: United States Pharmacopeial Convention: 2659 – 2663.

J. Nutrition and Food Processing

Copy rights @ Zakari David Adeiza.



This work is licensed under Creative Commons Attribution 4.0 License

To Submit Your Article Click Here:

Submit Manuscript

DOI:10.31579/2637-8914/165

## Ready to submit your research? Choose Auctores and benefit from:

- > fast, convenient online submission
- > rigorous peer review by experienced research in your field
- > rapid publication on acceptance
- > authors retain copyrights
- > unique DOI for all articles
- > immediate, unrestricted online access

At Auctores, research is always in progress.

Learn more <a href="https://auctoresonline.org/journals/nutrition-and-food-processing">https://auctoresonline.org/journals/nutrition-and-food-processing</a>

Auctores Publishing LLC – Volume 6(9)-165 www.auctoresonline.org ISSN: 2637-8914